## Lucifer Yellow Permeability Assay using Falcon<sup>®</sup> HTS 96 Multiwell Insert Systems

Protocol



For permeability studies, Caco-2 cell monolayers grown on Falcon HTS 96 Multiwell Insert Systems (Cat. Nos. 351130 or 351131) should be placed onto a Falcon 96 square well, angled-bottom plate (Cat. No. 353925) for analysis of lucifer yellow permeability. Although the lid and feeder tray of the Falcon HTS 96 Multiwell Insert System is non-directional, the insert plate is designed to be placed on the Falcon 96 square well, angled-bottom plate (Cat. No. 353925) in one unique orientation to prevent cross contamination of wells. To properly align the Falcon 96 Multiwell Insert Plate in the Falcon 96 square well, angled-bottom plate, make sure the Falcon logos on the top of both pieces face the same direction. The sampling ports on the insert plates face the same direction as the notched corner side of the 96 square well, angled-bottom plate.

Note: The standard Falcon 96 well plates are not compatible with the Falcon 96 Multiwell Insert System. Use of standard 96 well plates will result in media wicking up on the sides of the wells and possibly into the insert or out of the well.

### **Materials**

- Caco-2 cells grown on Falcon HTS 96 Multiwell Insert System, 1.0 μm pore size (Cat. Nos. 351130 or 351131)
- Falcon 96 square well, angled-bottom plate and lid (Cat. No. 353925)
- Lucifer Yellow (Molecular Probes)
- Transport buffer (HBSS with Ca<sup>2</sup>+, Mg<sup>2</sup>+, +10 mM HEPES, pH 7.4, phenol-red free)
- Platform orbital shaker
- Fluorescence plate reader

### **Lucifer Yellow Permeability Assay**

Transport buffer (HBSS with Ca<sup>2+</sup>, Mg<sup>2+</sup>, +10 mM HEPES, pH 7.4) is added to the basal compartment. Lucifer yellow is diluted in transport buffer and added to the apical compartment at a final concentration of 100  $\mu$ M. The monolayers are placed on a shaker at 70-90 rpm in a 37°C incubator with 90% relative humidity and 5% CO<sub>2</sub> for 1-2 hours. Fluorescence leakage was determined for lucifer yellow by 485 nm excitation and 530 nm emission using a fluorescence plate reader.

- a) Remove the Falcon 96 Multiwell Insert Plate from its feeder tray and place it directly on the Falcon 96 square well, angled-bottom plate (Cat. No. 353925, sold separately).
- b) Gently remove medium from each insert. Wash gently with transport buffer.
- c) Gently add 50  $\mu\text{L}$  of lucifer yellow dissolved in transport buffer (100  $\mu\text{M})$  to the inside of each insert.
- d) Add 250-275  $\mu$ L of transport buffer to each well of the Falcon 96 square well, angled-bottom plate.
- e) Incubate in a 37°C incubator (5% CO<sub>2</sub> and 90% humidity) for 1-2 hours on an orbital shaker set for 70-90 rpm.
- f) For a lucifer yellow standard curve, add increasing concentrations of lucifer yellow solution ranging from 0.1-50  $\mu$ M to a separate Falcon 96 square well, angled-bottom plate.

- g) Following incubation, remove the Falcon 96 Multiwell Insert Plate from the 96 square well, angledbottom plate and set aside. Lucifer yellow fluorescence in the 96 square well, angled-bottom plate (fluorescence leakage across the Caco-2 monolayer) is read directly in a fluorescence plate reader using a 485 nm excitation and an emission filter of 530 nm. The standard curve plate is also read directly in the fluorescence plate reader.
- h) Use the standard curve to calculate the lucifer yellow concentration in each well. These values can then be used to determine the % flux and permeability coefficients.

#### **Permeability Measurements**

The apical-to-basal permeability coefficients (Pc) can be calculated according to the following equation:

Pc = (V/(A x Ci)) x (Cf/T) where V is the volume of the basal chamber (mL), A is the area of the membrane insert (cm<sup>2</sup>), Ci is the initial concentration of the drug  $\mu$ M or fluorescence units/ml added), Cf is the final concentration of the drug ( $\mu$ M or fluorescence units/ml), and T is the assay time (seconds). Typical volumes include 50  $\mu$ L in apical compartment and 270  $\mu$ L transport buffer in basal compartment. Area of membrane = 0.0804 cm<sup>2</sup>.

#### Corning acquired the Falcon<sup>®</sup> brand.

**Corning Incorporated** 

Building 300, Suite 3401 Tewksbury, MA 01876 t 800.492.1110 t 978.442.2200 f 978.442.2476

www.corning.com/lifesciences

Life Sciences 836 North St. For additional Corning product, technical, or distributor information, please e-mail us at CLSTechServ@corning.com, visit our website www.corning.com/lifesciences or call 800.492.1110. Outside the United States, call 978.442.2200. For information on the acquisition, visit www.corning.com/discoverylabware.

Worldwide Support Offices ASIA/PACIFIC	Japan t 81 3-3586 1996 f 81 3-3586 1291 Korea	EUROPE France t 0800 916 882 f 0800 918 636	All Other European Countries t 31 (0) 20 659 60 f 31 (0) 20 659 76
Australia/New Zealand t 0402-794-347	t 82 2-796-9500 f 82 2-796-9300	Germany t 0800 101 1153 f 0800 101 2427 The Netherlands t 31 20 655 79 28 f 31 20 659 76 73	LATIN AMERICA Brasil t (55-11) 3089-741 f (55-11) 3167-070 Mexico t (52-81) 8158-840 f (52-81) 8313-858
China	Singapore		
t 86 21 2215 2888 f 86 21 6215 2988	t 65 6733-6511 f 65 6861-2913		
India	Taiwan		
t 91 124 4604000 f 91 124 4604099	t 886 2-2716-0338 f 886 2-2516-7500	<b>United Kingdom</b> t 0800 376 8660 f 0800 279 1117	

# CORNING | FALCON° CEllgro° Axyg=n. pyrex° Gosselin°

For Research Use Only. Not for use in diagnostic or therapeutic procedures.

For a listing of trademarks, visit us at www.corning.com/lifesciences/trademarks. Corning Incorporated, One Riverfront Plaza, Corning, NY 14831-0001